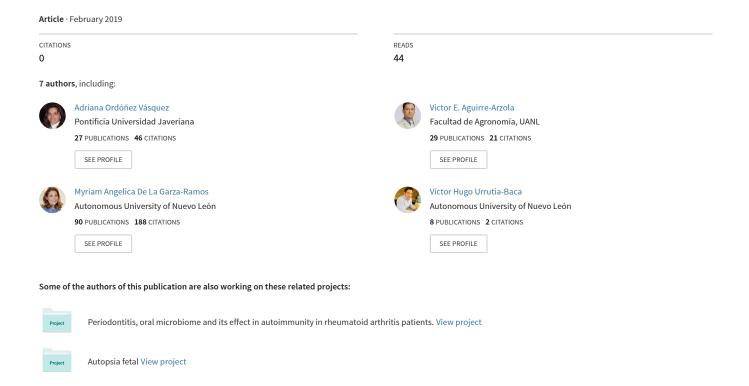
OPEN ACCESS International Journal of Pharmacology Review Article Toxicity, Teratogenicity and Anti-cancer Activity of α -solanine: A Perspective on Anti-cancer Potential



ISSN 1811-7775 DOI: 10.3923/ijp.2019.



Review Article

Toxicity, Teratogenicity and Anti-cancer Activity of α -solanine: A Perspective on Anti-cancer Potential

¹Adriana Ordóñez-Vásquez, ²Víctor Aguirre-Arzola, ³Myriam Angélica De la Garza-Ramos, ⁴Víctor Hugo Urrutia-Baca and ⁵Fernando Suárez-Obando

¹Institute of Human Genetics, School of Medicine, Pontifical Javeriana University, Cra. 7 No. 40-62, Building 32 Bogotá, Colombia ²School of Agronomy, Autonomous University of Nuevo, Francisco Villa s/n, Ex-Hacienda El Canada, Escobedo, N.L, Mexico ³Integral Dentistry and Specialties Unit, Research and Development Center in Health Sciences, Autonomous University of Nuevo León, School of Dentistry, Autonomous University of Nuevo León, Eduardo Aguirre Pequeño y Silao s/n, Mitras Centro, Monterrey, N.L., Mexico ⁴Integral Dentistry and Specialties Unit, Research and Development Center in Health Sciences, Autonomous University of Nuevo León, Eduardo Aguirre Pequeño y Silao s/n, Mitras Centro Monterrey, N.L., Mexico

⁵Institute of Human Genetics, School of Medicine, Pontifical Javeriana University, Cra. 7 No. 40-62, Building 32 Bogotá, Colombia, University Hospital San Ignacio, Cra. 7, No. 40-62, Building 24, Bogotá, Colombia

Abstract

The α -Solanine is a glycoalkaloid metabolite produced by solanaceae species, important plant food in the human nutrition. This α -solanine is highly toxic to animals and humans and has been indicated as a risk factor for developing congenital malformations. However, recent studies suggest that α -solanine possesses anti-microbial and anti-tumor activities. The aim of this review was to summarize the main properties of α -solanine, its toxicity and teratogenicity in animal models and the main findings reported about anti-cancer activity against various cancer cell lines in *in vitro* assays. Key α -solanine mechanisms of action are presented alongside arising interdisciplinary research, connecting agricultural sciences and medicine. Data presented in this review, may assisted in preventing toxic effects of α -solanine and promote research about its potential use in the treatment and management of human cancers.

Key words: α-Solanine, plant foods, teratogenicity, antitumor activities, glycoalkaloid metabolite, solanaceae species, anti-cancer activity

Received: Accepted: Published:

Citation: Adriana Ordóñez-Vásquez, Víctor Aguirre-Arzola, Myriam Angélica De la Garza-Ramos, Víctor Hugo Urrutia-Baca and Fernando Suárez-Obando, 2019. Toxicity, teratogenicity and anti-cancer activity of α-solanine: A perspective on anti-cancer potential. Int. J. Pharmacol., CC: CC-CC.

Corresponding Author: Myriam Angélica De la Garza-Ramos, Integral Dentistry and Specialties Unit, Research and Development Center in Health Sciences, Autonomous University of Nuevo León, School of Dentistry, Autonomous University of Nuevo León, Eduardo Aguirre Pequeño y Silao s/n, Mitras Centro, Monterrey, N.L., Mexico Tel: (+52) 8329-4000 Ext. 1816

Copyright: © 2019 Adriana Ordóñez-Vásquez et al. This is an open access article distributed under the terms of the creative commons attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Solanaceae plant family includes numerous species that are important for human nutrition, such as peppers, aubergines, tomatoes and varieties of potatoes. Solanaceae species produce alkaloids such as α -, β -, γ -solanine, α -, β -, γ -chaconine, solanidine and tomatidenol^{1,2}. The α -Solanine is a glycoalkaloid found mainly in potatoes (*Solanum tuberosum*) and other plant foods such as apples (*Malus domestica*), cherries (*Prunus avium*), eggplant (*Solanum melongena*) and tomatoes (*Solanum lycopersicum*). This glycoalkaloid was biochemically characterized in European black nightshade berries (*Solanum nigrum*) by Desfosses in 1820³. Since then, the concentration of α -solanine has been shown to vary in different plant parts, localizing in stems, leaves, husks and inside the tubers⁴, where this glycoalkaloid exerts a protective effect against fungi, bacteria and insects⁵.

The α -Solanine has been studied as a factor relevant for human health, mainly due to its toxicity and possible teratogenic effects in humans. Exposure to α-solanine has been linked to dozens of deaths in the 20th century in Germany and Britain⁶ and numerous cases of intoxication have been reported in various countries^{7,8}. Symptoms of α -solanine intoxication (respiratory distress, nausea, vomiting and diarrhea) are related to inhibition of acetylcholinesterase⁹, however, the underlying mechanisms of action are still under study. Teratogenicity of α -solanine has been demonstrated in murine and amphibian animal models, where it induced embryonic malformations, mainly of the central nervous system, such as exencephaly, encephalocele and anophthalmia¹⁰⁻¹². Recently, potential anti-cancer properties of α-solanine have received attention as it was shown to inhibit the growth of breast, pancreatic and melanoma cancer cells¹³⁻¹⁵.

As α -solanine is present in widely consumed human foods such as potato tubers, this review aimed to describe aspects of this alkaloid of relevance to human health, including effects on embryogenesis of the central nervous system. Importance of the quantification of α -solanine in the consumption tubercle is highlighted, inviting inter-disciplinary research between agricultural sciences and human medicine. Data presented in this review, may assist in preventing toxic effects of α -solanine and promote its use in the treatment and management of human cancers.

TOXICITY, TERATOGENICITY AND ANTI-CANCER OF α -SOLANINE

Solanaceae species produce a wide variety of nitrogencontaining secondary metabolites, including alkaloids¹⁶. Alkaloids play an essential role in protecting the plant against infection as they possess anti-microbial and insecticidal properties including inhibition of bacterial growth¹⁷, anti-fungal¹⁸, larvicidal and anti-oviposition activity¹⁸. Most alkaloids are toxic to animals, including man¹⁹.

Glycoalkaloid α-solanine is an organic, insoluble compound with a triterpenoid structure and is naturally found in plants of the Solanaceae family, mainly in stems, leaves, skin and inside the fruits. Mammals directly consume this compound in assorted dietary vegetables such as potatoes (Solanum tuberosum), tomatoes (Solanum lycopersicum), eggplant (Solanum melongena) and peppers (Capsicum annuum). Biochemically, α -solanine and other steroid alkaloids are derived from sterols, which are glycosylated to produce bitter-tasting steroidal glyco-alkaloids. Biosynthetic precursor of glyco-alkaloids is cholesterol. In plants, such as potatoes, cholesterol is cyclized to solanidine, which is subsequently glyco-sylated to α -solanine or α -chaconine²⁰. Together, these two compounds form up to 95% of total glyco-alkaloids in potatoes. External factors encountered during post-harvest handling of potatoes, such as exposure to light, heat, wounds and stress, significantly increase glyco-alkaloid content of the tubers, increasing the risk of post-consumption toxicity in animals and humans^{21,22}.

Among external factors affecting the concentration of glyco-alkaloids, exposure to ultraviolet (UV) light is the most studied^{23,24}. However, pathogenic factors such as infection by *Erwinia carotovora*, *Synchytrium endobioticum* and *Phytophthora infestans* are also relevant²⁵. Glyco-alkaloid content also depends on the variety of the potato, the geographical location of the crop and the conditions of storage, transport and marketing²⁶. Section of the potato tuber analyzed is also of importance as glyco-alkaloid concentration is lower in the meat than in green parts such as skin, eyes and shoots^{26,27}.

Several studies evaluated the concentration of glyco-alkaloids in different varieties of the tuber, analyzing crops in Pakistan^{26,27}, Denmark²⁷, Brazil²⁸, Ireland²⁹, Bolivia³⁰ and Canada³¹. Using high-performance liquid chromatography (HPLC) and reverse phase-based (RP-HPLC) techniques with UV detection (at 202 nm), these studies demonstrated that a typical tuber contains 12-20 mg kg⁻¹ of glyco-alkaloids, whereas a green tuber contains³² 250-280 mg kg⁻¹. Solanine concentration in commercial potato varieties should be and usually²⁸ is <200 mg kg⁻¹. However, some of the varieties analyzed in the afore-mentioned studies contained more than 200 mg kg⁻¹ glyco-alkaloids, which is the safe upper limit for human consumption recommended by the Food and Agriculture Organization (FAO) and the World Health Organization (WHO)^{26,28,33}. Finally, a synergic effect of the plant genotype and the environment was

observed on the glyco-alkaloid content of wild-type and commercial potato varieties²⁹.

Furthermore, quality of the tuber is determined, in part, by taste and color, parameters directly affected by the concentration of glyco-alkaloids because both bitter taste and green color are the result of glyco-alkaloid accumulation are insufficiently effective in reducing the glyco-alkaloid toxicity 32,35 as these compounds are thermostable, bioaccumulate and remain active after cooking and processing the potato 32 . However, the concentration of glyco-alkaloids decreases by 70% if potato skin is removed and by \leq 30% if the tuber is bleached or washed. Furthermore, glyco-alkaloid activity reduces by 92 and 83% at frying temperatures used for preparing French fries and pan-fried potatoes, respectively 36 .

Acute toxicity of \alpha-solanine: Acute α -solanine intoxication was characterized mainly by gastrointestinal symptoms (nausea, vomiting, diarrhea and gastrointestinal bleeding). In severe cases, a generalized rash is accompanied by neurological disorders, including cerebral edema, coma and death 19.

Outbreaks of α -solanine poisoning have been reported throughout the 20th century in several countries $^{37-39}$. The characteristic taste of potatoes was attributed to glyco-alkaloids, although these same compounds at high concentrations cause bitterness and a burning sensation in the mouth, which may prevent food poisoning 29 . Most potatoes purchased commercially contain low concentrations of α -solanine (4-10 mg/100 g dry weight) but if α -solanine concentration is \geq 20 mg/100 g, the tubers present a toxic hazard 40 . The maximum tolerated dose in humans is 1 mg kg $^{-1}$ body mass, whereas acute doses of 2-5 and 3-6 mg kg $^{-1}$ body mass had been reported as potentially lethal (Table 1) $^{29,39,41,42,10-12,43-48}$.

α-solanine teratogenicity: Potential teratogenic effects of glyco-alkaloids have been a cause of concern for public health since the 1970s, when specific congenital defects in humans, mainly neural tube defects (NTDs) were first associated with the ingestion of tubers producing these substances by Renwick et al.⁴⁹, based on a review of epidemiological data on an encephaly and spina bifida. Renwick et al.⁴⁹ postulated a link between the incidence of these neuronal dysplasias and consumption of potatoes stored during the winter and drawing from conclusions of previous studies, suggested that potatoes could become teratogenic as they get older or that a potato-infecting fungus could be a determining factor in the etiology of an encephaly and spina bifida. This hypothesis was based on overlapping geographical distributions of potato and NTDs⁵⁰. Since then, teratogenic effects of α-solanine in

the development of NTDs have been demonstrated in murine models⁴⁹ and in rabbits and pigs where this glyco-alkaloid caused defects in neurulation and anencephaly⁵¹. However, these teratogenic effects were not observed in non-human primates⁴⁷.

The estimated daily average consumption of potato glyco-alkaloids for a human is 12.75 mg (around 0.18 mg kg $^{-1}$ b.wt.), which is approximately 1/5th of the acute toxic concentration in humans (1 mg kg $^{-1}$ of b.wt.), making acute intoxication seemingly more relevant than a critical effect during embryo development. However as α -solanine is an insoluble substance that accumulates in the body up to 24 h after ingestion, the developing fetus could be affected by accumulation, not necessarily by acute exposure, making both the effects of acute intoxication and teratogenic impact of α -solanine accumulation worth considering. However, no existing evidence supported this hypothesis as teratogenic effects leading to NTDs manifest at critical points of development, not corresponding to a cumulative effect 52 .

In murine models, in utero exposure of embryos to α -solanine has consistently resulted in the development of central nervous system abnormalities, such as exencephaly, encephalocele and anophthalmia⁴². The NTDs are characterized by neurulation defect in which the closure of the anterior or posterior neuropore fails⁵³. It is hypothesized that the teratogenic threshold of α -solanine can be reached accumulatively or by exposure to increasing doses that impede cell differentiation, through alterations in cell morphology that inhibit aggregation. The α -solanine was shown to exhibit dose-dependent toxicity in rat stem cells, modifying cell morphology and causing decreased cell size and detachment of adherent cells in culture (decreased intercellular adhesion)⁵⁴, mechanisms that together could contribute to the closing defect of the neuropore.

In murine models, in addition to NTDs, fetal exposure to glyco-alkaloids such as α -solanine also resulted in alterations in the morphology of cardiac cells⁵⁵ and necrosis in the gastro-intestinal tract⁵⁶. In Xenopus embryos, α -chaconine displayed a more pronounced teratogenic effect than α -solanine⁴³, however, these glyco-alkaloids altered the embryonic development of Xenopus synergistically, with their combination shown to be the primary factor increasing the risk of developmental abnormalities¹². In fish embryos (*Oryzius latipes* or Japanese rice-fish and *Oncorhynchus mykiss* or rainbow trout), α -solanine increases mortality by inducing functional and structural defects of various organ systems. Additionally, *in vitro* exposure of cattle zygotes to α -solanine inhibits implantation and embryonic

		(IIIIII)	Physical properties of g-solanine ^{39,41}
	H _{II}	·	nrystate properties or a southing. Molecular formula: C45H73NO15 Molecular weight: 868,071 g moL ⁻¹
	/	.	Melting point: 285°C Water solubility: 1,380 mg L⁻¹ (25°C)
	HIIII	п	PubChem CID: 129627772
	Luft.		
	Н	ш	
		Onter	
	H O H	H,OH	
		严	
	H H		
Toxicology	H Safe potato concentration	Safe oral dose (humans)	Potential lethality
V	<200 mg kg ⁻¹ in fresh potato	1 mg kg ⁻¹ body mass	2-5 mg kg ⁻¹ body mass to 3-6 mg kg ⁻¹ body mass
Teratology A	Animal model Syrian hamsters	System affected Central nervous system	Phenotype Exencephaly, encephalocele and anophthalmia ⁴²
U ×	Chick embryos Xenonis	Central nervous system, cardiac system, somite development Central nervous system, dastrointestinal system	*Cranioschisis, caudal regression syndrome, cardiac septal defects ^{10,11} ***Microrenhalv anencenhalv ahnormal cuit coilinoi ^{12,43}
, -	Japanese rice fish (Oryzius lutipes)		Increased embryo mortality at high concentrations ⁴⁴
± ∗ C	***Pig (oocyte and embryo)	Oocyte maturation, embryonic development	Increased embryo mortality at high concentrations** Disturbed meiosis, reduced cleavage and blastocyst formation, decreased number of total and inner cell mass cells*
ш	Bovine (Oocyte and emblyo) Female rhesus monkeys and marmosets Aafora braading and 6 weeks following		nminitori or pre-mipantation embryo ueverlopment, reduced monta creavage rates. Absence of neural tube defects, development of hydrocephalus in rhesus monkey infants ⁰
· ن ر	conception)		
7	Drosophila melanogaster	Body size, thorax and abdomen	Drosophila melanogaster Body size, thorax and abdomen Decreased body size, deformed wings, smaller abdominal zone ⁴⁸

4

Table 1: Physical properties of $\alpha\text{-solanine, toxic}$ doses and teratology animal models

development⁴⁶. In contrast, glyco-alkaloids derived from *Solanum lycocarpum* (wolf apple) act as endocrinological disruptors^{57,58}.

This evidence indicated that developmental effects of α -solanine occur under particular circumstances, implying an all-or-nothing effect, similar to other drugs with teratogenic potential⁵⁹. Impact on higher primates could be severe but clinically undetectable as NTDs if exposure occurs during early developmental stages and causes a premature loss of the embryo, whereas if the teratogenic threshold is not reached, development would not be significantly altered and would continue un-obstructed.

Increased consumption of foods with high folic acid content was shown to be an effective measure for preventing NTDs^{60,61}. As folic acid inhibits the growth of pathogenic bacteria in potatoes such as *Erwinia* spp.⁶², it has been suggested that bio-fortification of potato with folic acid would have a triple effect: Preventing potato diseases by inhibiting pathogen growth⁶², decreasing α -solanine concentration as less alkaloid would be required to defend the tuber against pathogens and decreasing teratogenic potential⁵³. Table 1 summarized teratologic effects of α -solanine in diverse animal models.

α-solanine and cancer: Alongside protective actions in the plant and potential teratogenic effects in animals, α-solanine has demonstrated *in vitro* and *in vivo* anti-cancer activity, with most studies identifying activation of apoptosis as the underlying mechanism of α-solanine anti-tumor activity⁶³.

Apoptotic effects of α -solanine have been demonstrated in vitro in HepG2 cells², in which α -solanine caused cell cycle arrest, decreased the duration of G phase² and increased the duration of S phase of the cell cycle as well as decreased the synthesis of anti-apoptotic regulatory protein⁶⁴ Bcl-2. Additionally, α-solanine induced HepG2 cellular morphological changes typical of apoptosis, inducing changes in the mitochondrial membrane potential, altered calcium gradients⁶⁵ and increased synthesis of reactive oxygen species (·OH and H₂O₂)⁶⁶. Likewise, α-solanine decreased the synthesis of histone deacetylase 1 (HDAC1), which regulates cell growth, while stimulating the synthesis of apoptosis-inducing proteins ASK1 (apoptosis signal-regulating kinase 1) and TBP-2 (tetrahymena piggyBac transposase 2)66, leading to decreased cell proliferation and increased rate of programmed cell death.

In models of colon cancer, induction of apoptosis is mediated mainly by activation of caspase-3 pathways and inhibition of phosphorylation of ERK1 and ERK2 (extracellular signal-regulated protein kinases 1 and 2)⁶⁷. Caspase-3 is a

pro-apoptotic kinase⁶⁸, whereas ERK1 and ERK2 participate in diverse cellular functions, including cell cycle progression, migration, survival, differentiation, metabolism, proliferation and transcription. In these models, exposure to α -solanine favored programmed cell death⁶⁹. Additionally, increased rate of autophagy⁷⁰ was observed in pancreatic cancer cells exposed to α -solanine, in which the alkaloid suppressed the Akt/mTOR pathway (phosphatidylinositol 3-kinase/Akt/mammalian target of rapamycin)⁶³, involved in proliferation of diverse cancer cell types⁷¹.

In prostate cancer, in both *in vitro* models and *in vivo* mouse models, α -solanine displayed apoptotic effects mediated by synergistic cyclin suppression, induction of reactive oxygen species and activation of P38. P38 is a protein belonging to the sub-family MAPK (mitogen-activated protein kinases)⁷² that, similar to ERK1/2 proteins, regulates cell cycle, proliferation and intercellular interactions⁷³.

Neo-angiogenesis and vascular proliferation are fundamental mechanisms in tumor survival. The α -chaconine inhibits proliferation of endothelial cells by reducing the expression of MMP-2 (matrix metalloproteinase-2), a protein involved in angiogenesis⁷⁴. Similarly, α -solanine intervenes in vascular remodeling, reversing the effects of AXIN (axis inhibition protein 1) and BMPR2 (bone morphogenetic protein receptor type-2) proteins involved in tumor proliferation and metastasis through vascular remodeling mediated by β -catenin⁷⁵.

More than one mechanism, triggered by induction of apoptosis may explain the effect of α -solanine on tumor proliferation as exemplified by α -solanine activity in melanoma cells, in which cell migration is inhibited by decreased activity of MMP-2, JNK (c-Jun N-terminal kinase) and PI3K/Akt, implying a mechanism combining increase in apoptosis and regulation of angiogenesis¹⁴. Similar mechanisms of action have been described in models of breast cancer⁵. Additionally, in lung adenocarcinoma and esophageal cancer cells, exposure to α -solanine increased radiosensitivity and expression of microRNA-13876,77. In conclusion, α-solanine shows promising pharmacological potential for the management of human cancers by stimulating apoptosis, inhibiting angiogenesis, regulating the cell cycle and increasing cellular sensitivity to radiotherapy. The Table 2 summarized physiological effects of α -solanine in different tissues and human diseases associated with animal models and *in vitro* assays analyzed⁷⁸⁻⁸³, whereas Table 3 summarized the effect of α -solanine in different cancers^{72,84,85,5,15,76,13,14,66}

Current results demonstrated that α -solanine produces structural changes and growth inhibition in Mensequimal

200	200
dicas	200
200	5
3	3
4	200
ij	5
20	222
۳	2
'n	5
9	3
2	2
4	3
9	5
ż	Ē
2	ş
ģ	٥
ď	5
145	
2	=
9	2
.ic	5
2	5
خ	5
9	2
j	2
Ho I	5
	3
5	5
,ioi	Š
P	Ē
Ċ	,
H	3
Ë	Ξ

System	Model	Physiological effects	Related disease	Genetic background
Vascular system	Monocrotaline-induced pulmonary Reverse	Reversed pulmonary vascular	Pulmonary arterial hypertension	α-solanine reversed dysfunctional AXIN2, β-catenin and BMPR2 signaling
		remodeling and vascular angiogenesis78		
Gastrointestinai tract	Cultured epitnellal cell lines of rat increase and human intestinal mucosa	Increased brush border permeability?	Inflammatory bowel disease	
	Murine small intestine Disrupti	Disruption of intestinal barrier integrity ⁸⁰	Inflammatory bowel disease	
	Caco-2 monolayers Cytotox tight jur	Cytotoxicity and disruption of intestinal tight junction integrity ⁸¹	Intestinal autoimmune diseases	
Immune system	Plasmodium yoelii 17XL	α-chaconine showed a dose-dependent		
	infection in mice suppres	suppression of malaria infection. Simultaneous		
	adminis did not	administration of α -chaconine and α -solanine did not show any synergistic effects 82	Malaria	
Immune system	LPS-induced septicshock in mice	Inhibition of the NF-kB signaling pathway83	Endotoxin-induced shock	Inhibition of LPS-activation of nuclear factor-xB (NF-xB) reduced translocation
				of p65, degradation of inhibitory $\kappa B\alpha$ (IkB α) and phosphorylation of IkB kinase α/β (IKK α/β)
Table 3: α-solanine and cancer	cancer			
Cancer type	Experimental model	α-solanine mechanisms of action		
Prostate cancer	Cultured human prostate cancer cell line DU145	Inhibition of prostate cancer grow	h by blocking the expression of cell cycl	Inhibition of prostate cancer growth by blocking the expression of cell cycle proteins (cyclin D1, cyclin E1, CDK2, CDK4, CDK6 and P21) and inducing
		apoptosis via reactive oxygen spec	apoptosis via reactive oxygen species and activation of P38 pathway'²	
	Cultured human prostate cancer cell line PC-3	Inhibition of proliferation and indu	ction of apoptosis of tumor cells by redu	Inhibition of proliferation and induction of apoptosis of tumor cells by reducing mRNA levels of MMP-2, MMP-9 and extracellular inducer of matrix
		metalloproteinase (EMMPRIN) and increasing the exp of metallo-proteinases 1 and 2 (TIMP-1 and TIMP-2) 84	increasing the expression of reversion-ii AP-1 and TIMP-2) ⁸⁴	metalloproteinase (EMMPRIN) and increasing the expression of reversion-inducing cysteine-rich protein with Kazal motifs (RECK) and tissue inhibitors of metallo-proteinases 1 and 2 (TIMP-1 and TIMP-2)**
Esophageal carcinoma	Cultured human esophageal EC9706/Eca109 cancer cells		MMP-9 and increased apoptosis85	
Breast cancer	Mice breast cancer	Decreased expression of anti-apoptotic Bd-2 protein and increased Reduced expression levels of Bax, MMP-2, MMP-9, mTOR and Akt ¹⁵	totic Bcl-2 protein and increased expres MMP-2, MMP-9, mTOR and Akt ¹⁵	Decreased expression of anti-apoptotic Bcl-2 protein and increased expression of Bcl-2-like protein 4 (proapoptoticBax protein) ⁵ Reduced expression levels of Bax, MMP-2, MMP-9, mTOR and Akt ¹⁵
Lung adenocarcinoma	Cultured A549 and H1299 cells	Inhibited cell migration and invasio	inhibited cell migration and invasion ability and induced expression of miR-138 76	-13876
Pancreatic cancer	Cultured human pancreatic cancer cells (PANC-1 cell line)		s proliferation by decreased expression c	Inhibition of pancreatic cancer cells proliferation by decreased expression of VEGF and suppressed mRNA expression of MMP-2, MMP-9, ENOS, EMMPRIN, and CD44 ¹³
Melanoma	Human melanoma cell line A2058	Suppressed phosphorylation of JNK, PI3K and Akt14	K, PI3K and Akt ¹⁴	
Liver cancer	HepG2 cells	Increased expression of ASK1 and TE	3P-2, induced ROS production and inhibite	Increased expression of ASK1 and TBP-2, induced ROS production and inhibited expression of proliferation-associated proteins such as HDAC1, all contributing
		lo elevateu apoptosis tate		

stem cells (MSCs) in a dose-dependent manner, consistent with findings in other cell types, such as, colon (HT29, T84) and liver (HepG2) cancer cells⁸⁶. In addition, an important finding obtained in present study is the fact that α -solanine can affect the adhesion capacity of MSCs as the dose increases, a phenomenon that can be directly associated with the reported morphological alterations.

Perspectives: Further evidence of potential teratogenic effects of α -solanine in humans is required before specific public health recommendations can be issued, however, as α -solanine has shown teratogenic effects in various animal models, it should not be ruled out as a risk factor for the development of congenital malformations, especially NTDs. α -solanine is a versatile substance, which, in addition to protecting the plant that produces it, shows anti-tumor activity and taking into account its effects on angiogenesis, may be beneficial in treating vascular disorders such as primary pulmonary hypertension⁷⁷.

Additionally, continued research of biological effects exerted by α -solanine in normal and cancer cells should establish mechanisms involved in toxicological and teratogenic effects of α -solanine and make further progress in preventing cancer progression. As natural sources of α -solanine are potatoes and other widely consumed solanaceae plants, research in this area is an opportunity for inter-disciplinary studies combining agricultural sciences and medicine. It should be noted that glyco-alkaloids are also found as a highly valuable raw material in the residues of the potato industry.

High variability of α -solanine concentration in potatoes and high tuber consumption rates highlight importance of determining specific glyco-alkaloid concentrations in all native varieties of the tuber, different commercial forms (frozen, fried or dehydrated potato) and in new and improved potato varieties currently developed. Altitude, climate, storage and geographical location should be taken into account and levels of α-solanine correlated with diverse biological and environmental conditions. These recommendations become relevant in the light of the precautionary principle, which implies that strategies must be established to contain risks that to date do not have a complete scientific understanding^{78,87}. Terato-genic potential of α -solanine falls under this principle as only indirect evidence of harmful effects of α-solanine on human embryonic development has been discovered.

CONCLUSION

The α -solanine is a glyco-alkaloid metabolite produced by Solanaceae species, mainly in potatoes, important plant foods in the human nutrition. It has been reported the α -solanine teratogenicity and toxicity in *in vitro* assays and *in vivo* animal model. Accordingly, further studies of α -solanine in humans to prevent harmful effects are needed. Research in this area could be an opportunity for inter-disciplinary studies that combine agricultural sciences and medicine. In addition, the continuous investigation of the biological effects exerted by α -solanine in normal and cancerous cells could establish the molecular mechanisms involved before promoting its possible use in the treatment and control of management cancer.

SIGNIFICANCE STATEMENT

This study discover and summarize the biological effects reported up to now of α -solanine and its anti-cancer potential. α -solanine is a metabolite produced by Solanaceae species, important plant foods of human consumption. This review article can be beneficial in the formation of a general concept about toxic and terato-genic effects of α -solanine to take into consideration in human nutrition. This study will help the researcher to uncover the critical areas of research about its potential use in the treatment and management of human cancers that many researchers were not able to explore. Thus a new theory on the beneficial properties of α -solanine may be arrived at.

ACKNOWLEDGMENT

We would like to thank Editage (www.editage.com) for English language editing.

REFERENCES

- Schreiber, K., 1968. Steroid Alkaloids: The Solanum Group. In: The Alkaloids: Chemistry and Physiology, Manske, R.H.F. (Ed.)., Academic Press, USA., pp: 1-192.
- Friedman, M., 2015. Chemistry and anticarcinogenic mechanisms of glyco-alkaloids produced by eggplants, potatoes and tomatoes. J. Agric. Food Chem., 63: 3323-3337.
- 3. Conner, H.W., 1937. Effect of light on solanine synthesis in the potato tuber. Plant Physiol., 12: 79-98.
- Matsuda, F., K. Morino, H. Miyazawa, M. Miyashita and H. Miyagawa, 2004. Determination of potato glyco-alkaloids using high pressure liquid chromatography–electrospray ionisation/mass spectrometry. Phytochem. Anal.: Int. J. Plant Chem. Biochem. Tech., 15: 121-124.

- Mohsenikia, M., A.M. Alizadeh, S. Khodayari, H. Khodayari and A. Karimi *et al.*, 2013. The protective and therapeutic effects of alpha-solanine on mice breast cancer. Eur. J. Pharmacol., 718: 1-9.
- Anonymous, 1979. Solanine poisoning. Br. Med. J., 2: 1458-1459.
- McMillan, M. and J.C. Thompson, 1979. An outbreak of suspected solanine poisoning in schoolboys: Examination of criteria of solanine poisoning. QJM: Int. J. Med., 48: 227-243.
- 8. Friedman, M. and R. Rasooly, 2013. Review of the inhibition of biological activities of food-related selected toxins by natural compounds. Toxins, 5: 743-775.
- Dolan, L.C., R.A. Matulka and G.A. Burdock, 2010. Naturally occurring food toxins. Toxins, 2: 2289-2332.
- Mun, A.M., E.S. Barden, J.M. Wilson and J.M. Hogan, 1975.
 Teratogenic effects in early chick embryos of solanine and glyco-alkaloids from potatoes infected with late blight, *Phytophthora infestans*. Teratology, 11: 73-77.
- 11. Jelinek, R., V. Kyzlink and C. Blattny, Jr., 1976. An evaluation of the embryotoxic effects of blighted potatoes on chicken embryos. Teratology, 14: 335-342.
- 12. Rayburn, J.R., M. Friedman and J.A. Bantle, 1995. Synergistic interaction of glyco-alkaloids α -chaconine and α -solanine on developmental toxicity in *Xenopus embryos*. Food Chem. Toxicol., 33: 1013-1019.
- 13. Lv, C., H. Kong, G. Dong, L. Liu and K. Tong *et al.*, 2014. Antitumor efficacy of α -solanine against pancreatic cancer *in vitro* and *in vivo*. Plos One, Vol. 9. 10.1371/journal. pone. 0087868.
- 14. Lu, M.K., Y.W. Shih, T.T.C. Chien, L.H. Fang, H.C. Huang and P.S. Chen, 2010. α -Solanine inhibits human melanoma cell migration and invasion by reducing matrix metalloproteinase-2/9 activities. Biol. Pharm. Bull., 33: 1685-1691.
- 15. Mohsenikia, M., B. Farhangi, A.M. Alizadeh, H. Khodayari and S. Khodayari *et al.*, 2016. Therapeutic effects of dendrosomal solanine on a metastatic breast tumor. Life Sci., 148: 260-267.
- 16. Krits, P., E. Fogelman and I. Ginzberg, 2007. Potato steroidal glyco-alkaloid levels and the expression of key isoprenoid metabolic genes. Planta, 227: 143-150.
- 17. Kumar, P., B. Sharma and N. Bakshi., 2009. Biological activity of alkaloids from *Solanum dulcamara* L. Nat. Prod. Res., 23: 719-723.
- Sanchez Maldonado, A.F., A. Schieber and M.G. Ganzle, 2016.
 Antifungal activity of secondary plant metabolites from potatoes (*Solanum tuberosum* L.): Glyco-alkaloids and phenolic acids show synergistic effects. J. Applied Microbiol., 120: 955-965.
- 19. Dalvi, R.R. and W.C. Bowie, 1983. Toxicology of solanine: An overview. Vet. Hum. Toxicol., 25: 13-15.

- 20. Petersson, E.V., U. Arif, V. Schulzova, V. Krtkova and J. Hajslova *et al.*, 2013. Glyco-alkaloid and calystegine levels in table potato cultivars subjected to wounding, light and heat treatments. J. Agric. Food Chem., 61: 5893-5902.
- 21. Friedman, M., J.N. Roitman and N. Kozukue, 2003. Glyco-alkaloid and calystegine contents of eight potato cultivars. J. Agric. Food Chem., 51: 2964-2973.
- 22. Friedman, M., 2006. Potato glyco-alkaloids and metabolites: Roles in the plant and in the diet. J Agric. Food Chem., 54: 8655-8681.
- 23. Phillips, B.J., J.A. Hughes, J.C. Phillips, D.G. Walters, D. Anderson and C.S.M. Tahourdin, 1996. A study of the toxic hazard that might be associated with the consumption of green potato tops. Food Chem. Toxicol., 34: 439-448.
- 24. Patil, B.C., O.K. Salunkhe and B. Singh, 1971. Metabolism of solanine and chlorophyll in potato tubers as affected by light and specific chemicals. J. Food Sci., 36: 474-476.
- 25. Morrissey, J.P. and A.E. Osbourn, 1999. Fungal resistance to plant antibiotics as a mechanism of pathogenesis. Microbiol. Mol. Biol. Rev., 63: 708-724.
- 26. Aziz, A., M.A. Randhawa, M.S. Butt, A. Asghar, M. Yasin and T. Shibamoto, 2012. Glyco-alkaloids (α chaconine and α-solanine) contents of selected Pakistani potato cultivars and their dietary intake assessment. J. Food Sci., 77: T58-T61.
- 27. Knuthsen, P., U. Jensen, B. Schmidt and I.K. Larsen, 2009. Glyco-alkaloids in potatoes: Content of glyco-alkaloids in potatoes for consumption. J. Food Compos. Anal., 22: 577-581.
- 28. Machado, R.M., M.C.F. Toledo and L.C. Garcia, 2007. Effect of light and temperature on the formation of glyco-alkaloids in potato tubers. Food Control, 18: 503-508.
- 29. Valcarcel, J., K. Reilly, M. Gaffney and N. O'Brien, 2014. Effect of genotype and environment on the glyco-alkaloid content of rare, heritage and commercial potato varieties. J. Food Sci., 79: T1039-T1048.
- Carrasco, E., N. Estrada, J. Gabriel, G. Alfaro and Y. Larondelle, 2016. Seis cultivares potenciales de papa con resistencia al tizon tardio (*Phytophthora infestans*) en Bolivia. Rev. Latinoam. Papa, 9: 106-122.
- 31. Ji, X., L. Rivers, Z. Zielinski, M. Xu and E. MacDougall *et al.*, 2012. Quantitative analysis of phenolic components and glyco-alkaloids from 20 potato clones and in vitro evaluation of antioxidant, cholesterol uptake and neuroprotective activities. Food Chem., 133: 1177-1187.
- 32. Omayio, D.G., G.O. Abong and M.W. Okoth, 2016. A review of occurrence of glyco-alkaloids in potato and potato products. Curr. Res. Nutr. Food Sci. J., 4: 195-202.
- 33. Langkilde, S., M. Schroder, T. Frank, L.V. Shepherd and S. Conner *et al.*, 2012. Compositional and toxicological analysis of a GM potato line with reduced α-solanine contenta 90-day feeding study in the Syrian Golden hamster. Regulat. Toxicol. Pharmacol., 64: 177-185.

- 34. Sanford, L.L. and S.L. Sinden, 1972. Inheritance of potato glyco-alkaloids. Am. Potato J., 49: 209-217.
- 35. Campos, S., C. Silva, P. Campana and V. Almeida, 2016. Toxicidade de especies vegetais. Rev. Bras. Plantas Med., 18: 373-378.
- 36. Tian, J., J. Chen, X. Ye and S. Chen, 2016. Health benefits of the potato affected by domestic cooking: A review. Food Chem., 202: 165-175.
- 37. Alexander, R.F., G.B. Forbes and E.S. Hawkins, 1948. A fatal case of solanine poisoning. Br. Med. J., Vol. 2.
- 38. Toda, M., C. Uneyama and F. Kasuga, 2014. [Trends of plant toxin food poisonings during the past 50 years in Japan]. Shokuhin Eiseigaku Zasshi, 55: 55-63.
- 39. Barceloux, D.G., 2009. Potatoes, tomatoes and solanine toxicity (*Solanum tuberosum* L., *Solanum lycopersicum* L.). Dis. Month, 55: 391-402.
- 40. Lee, M.R., 2006. The Solanaceae: Foods and poisons. J. R. College Phys. Edinburgh, 36: 162-169.
- 41. Kim, S., P.A. Thiessen, E.E. Bolton, J. Chen and G. Fu *et al.*, 2015. PubChem substance and compound databases. Nucleic Acids Res., 44: D1202-D1213.
- 42. Gaffield, W. and R.F. Keeler, 1996. Induction of terata in hamsters by solanidane alkaloids derived from *Solanum tuberosum*. Chem. Res. Toxicol., 9: 426-433.
- 43. Friedman, M., J.R. Rayburn and J.A. Bantle, 1991. Developmental toxicology of potato alkaloids in the frog embryo teratogenesis assay-Xenopus (FETAX). Food Chem. Toxicol., 29: 537-547.
- 44. Crawford, L. and R.M. Kocan, 1993. Steroidal alkaloid toxicity to fish embryos. Toxicol. Lett., 66: 175-181.
- 45. Lin, T., R.K. Oqani, J.E. Lee, J.W. Kang and S.Y. Kim *et al.*, 2018. α -Solanine impairs oocyte maturation and quality by inducing autophagy and apoptosis and changing histone modifications in a pig model. Reprod. Toxicol., 75: 96-109.
- Wang, S., K.E. Panter, W. Gaffield, R.C. Evans and T.D. Bunch, 2005. Effects of steroidal glyco-alkaloids from potatoes (*Solanum tuberosum*) on in vitro bovine embryo development. Anim. Reprod. Sci., 85: 243-250.
- 47. Allen, J.R., R.J. Marlar, C.F. Chesney, J.P. Helgeson and A. Kelman *et al.*, 1977. Teratogenicity studies on late blighted potatoes in nonhuman primates (*Macaca mulatta* and *Saguinus labiatus*). Teratology, 15: 17-23.
- Ventrella, E., Z. Adamski, E. Chudzinska, M. Miądowicz-Kobielska et al., 2016. Solanum tuberosum and Lycopersicon esculentum leaf extracts and single metabolites affect development and reproduction of Drosophila melanogaster. Plos One, Vol. 11. 10.1371/ journal.pone.0155958.
- Renwick, J.H., W.D.B. Claringbold, M.E. Earthy, J.D. Few, A. Carolines and S. McLean, 1984. Neural tube defects produced in Syrian hamsters by potato glyco-alkaloids. Teratology, 30: 371-381.

- 50. Renwick, J.H., A.M. Possamai and M.R. Munday, 1974. Potatoes and spina bifida. Proc. R. Soc. Med., 67: 360-364.
- 51. Sharma, R.P., C.C. Willhite, M.T. Wu and D.K. Salunkhe, 1978. Teratogenic potential of blighted potato concentrate in rabbits, hamsters and miniature swine. Teratology, 18: 55-61.
- 52. Harvey, M.H., B.A. Morris, M. McMillan and V. Marks, 1986. Potato steroidal alkaloids and neural tube defects: Serum concentrations fail to demonstrate a causal relation. Hum. Toxicol., 5: 249-253.
- 53. Suarez-Obando, F., A. Ordonez-Vasquez and I. Zarante, 2010. Defectos del tubo neural y acido folico: Patogenia, metabolismo y desarrollo embriologico. Rev. Literat., Rev. Colomb. Obstetr. Ginecol., 61: 49-60.
- 54. Ordonez-Vasquez, A., L. Jaramillo-Gomez, C. Duran-Correa, E. Escamilla-Garcia, D. la Garza-Ramos, M. Angelica and F. Suarez-Obando, 2017. A reliable and reproducible model for assessing the effect of different concentrations of α-Solanine on rat bone marrow mesenchymal stem cells. Bone Marrow Res., Vol. 2017. 10.1155/2017/2170306.
- 55. Bergers, W.W. and G.M. Alink, 1980. Toxic effect of the glycoalkaloids solanine and tomatine on cultured neonatal rat heart cells. Toxicol. Lett., 6: 29-32.
- 56. Baker, D.C., R.F. Keeler and W.P. Gaffield, 1988. Mechanism of death in Syrian hamsters gavaged potato sprout material. Toxicol. Pathol., 16: 333-339.
- 57. Schwarz, A., E.C. Felippe, M.M. Bernardi and H.S. Spinosa, 2005. Impaired female sexual behavior of rat offspring exposed to *Solanum lycocarpum* unripe fruits during gestation and lactation: Lack of hormonal and fertility alterations. Pharmacol. Biochem. Behav., 81: 928-934.
- 58. Nelson, J.E., 1992. Developmental staging in a marsupial *Dasyurus hallucatus*. Anat. Embryol., 185: 335-354.
- 59. Adam, M.P., 2012. The all or none phenomenon revisited. Birth Defects Res. Part A: Clin. Mol. Teratol., 94: 664-669.
- Vasquez, A.O. and F. Suarez-Obando, 2015. Neural tube defects and folic acid: A historical overview of a highly successful preventive intervention. Historia Ciencias Saude-Manguinhos, 22: 1157-1172.
- 61. Ordonez, A. and F. Suarez, 2006. Ascertaining a Colombian female university student population's knowledge about folic acid and its benefits for their reproductive health. Rev. Colombiana Obstetr. Ginecol., 57: 271-278.
- 62. Marcelo-Correa, A., A. Ordonez-Vasquez, A.A. Trespalacios and F. Suarez-Obando, 2017. Inhibicion del crecimiento de Erwinia chrysanthemi a diferentes concentraciones de acido folico: Posible uso del acido folico como agente bacteriostatico y fortificante de la papa *Solanum tuberosum*. Univ. Salud, 19: 140-148.
- 63. Hasanain, M., A. Bhattacharjee, P. Pandey, R. Ashraf and N. Singh *et al.*, 2015. α-Solanine induces ROS-mediated autophagy through activation of endoplasmic reticulum stress and inhibition of Akt/mTOR pathway. Cell Death Dis., Vol. 6. 10.1038/cddis.2015.219.

- 64. Ji, Y.B., S.Y. Gao, C.F. Ji and X. Zou, 2008. Induction of apoptosis in HepG₂ cells by solanine and Bcl-2 protein. J. Ethnopharmacol., 115: 194-202.
- 65. Gao, S.Y., Q.J. Wang and Y.B. Ji, 2006. Effect of solanine on the membrane potential of mitochondria in HepG₂cells and [Ca²+] i in the cells. World J. Gastroenterol., 12: 3359-3367.
- 66. Meng, X.Q., W. Zhang, F. Zhang, S.Y. Yin, H.Y. Xie, L. Zhou and S.S. Zheng, 2016. Solanine-induced reactive oxygen species inhibit the growth of human hepatocellular carcinoma HepG₂ cells. Oncol. Lett., 11: 2145-2151.
- 67. Yang, S.A., S.H. Paek, N. Kozukue, K.R. Lee and J.A. Kim, 2006. α-Chaconine, a potato glyco-alkaloid, induces apoptosis of HT-29 human colon cancer cells through caspase-3 activation and inhibition of ERK 1/2 phosphorylation. Food Chem. Toxicol., 44: 839-849.
- 68. Julien, O. and J.A. Wells, 2017. Caspases and their substrates. Cell Death Different., 24: 1380-1389.
- 69. Roskoski, Jr., R., 2012. ERK1/2 MAP kinases: Structure, function and regulation. Pharmacol. Res., 66: 105-143.
- 70. Ngabire, D. and G.D. Kim, 2017. Autophagy and inflammatory response in the tumor microenvironment. Int. J. Mol. Sci., Vol. 18. 10.3390/ijms18092016.
- 71. Barrett, D., V.I. Brown, S.A. Grupp and D.T. Teachey, 2012. Targeting the PI3K/AKT/mTOR signaling axis in children with hematologic malignancies. Pediat. Drugs, 14: 299-316.
- 72. Pan, B., W. Zhong, Z. Deng, C. Lai and J. Chu *et al.*, 2016. Inhibition of prostate cancer growth by solanine requires the suppression of cell cycle proteins and the activation of ROS/P38 signaling pathway. Cancer Med., 5: 3214-3222.
- 73. Cargnello, M. and P.P. Roux, 2011. Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. Microbiol. Mol. Biol. R., 75: 50-83.
- 74. Lu, M.K., P.H. Chen, Y.W. Shih, Y.T. Chang, E.T. Huang, C.R. Liu and P.S. Chen, 2010. alpha-Chaconine inhibits angiogenesis *in vitro* by reducing matrix metalloproteinase-2. Biol. Pharm. Bull., 33: 622-630.
- 75. Yang, H., C. Liu, R.M. Zhou, J. Yao and X.M. Li *et al.*, 2016. Piezo2 protein: A novel regulator of tumor angiogenesis and hyperpermeability. Oncotarget, 7: 44630-44643.
- 76. Zhang, F., R. Yang, G. Zhang, R. Cheng and Y. Bai *et al.*, 2016. Anticancer function of α -solanine in lung adenocarcinoma cells by inducing microRNA-138 expression. Tumor Biol., 37: 6437-6446.

- 77. Wang, Y., J. Wu, W. Guo, Q. Sun and X. Chen *et al.*, 2016. α -solanine modulates the radiosensitivity of esophageal cancer cells by inducing microRNA 138 expression. Cell. Physiol. Biochem., 39: 996-1010.
- 78. Nie, X., Y. Dai, J. Tan, Y. Chen and G. Qin *et al.*, 2017. α -Solanine reverses pulmonary vascular remodeling and vascular angiogenesis in experimental pulmonary artery hypertension. J. Hypertens., 35: 2419-2435.
- 79. Gee, J.M., G.M. Wortley, I.T. Johnson, K.R. Price, A.A.J.J.L. Rutten, G.F. Houben and A.H. Penninks, 1996. Effects of saponins and glyco-alkaloids on the permeability and viability of mammalian intestinal cells and on the integrity of tissue preparations *in vitro*. Toxicol. In Vitro, 10: 117-128.
- 80. Patel, B., R. Schutte, P. Sporns, J. Doyle, L. Jewel and R.N. Fedorak, 2002. Potato glyco-alkaloids adversely affect intestinal permeability and aggravate inflammatory bowel disease. Inflamm. Bowel Dis., 8: 340-346.
- 81. Glynn, A., A.M. Igra, S. Sand, N.G. Ilback, K.E. Hellenas, J. Rosen and B. Aspenstrom-Fagerlund, 2017. Are additive effects of dietary surfactants on intestinal tight junction integrity an overlooked human health risk?-A mixture study on Caco-2 monolayers. Food Chem. Toxicol., 106: 314-323.
- 82. Chen, Y., S. Li, F. Sun, H. Han and X. Zhang *et al.*, 2010. *In vivo* antimalarial activities of glyco-alkaloids isolated from Solanaceae plants. Pharm. Biol., 48: 1018-1024.
- 83. Shin, J.S., K.G. Lee, H.H. Lee, H.J. Lee and H.J. An *et al.*, 2016. α Solanine isolated from *Solanum tuberosum* L. cv jayoung abrogates LPS-induced inflammatory responses Via NF κ B inactivation in RAW 264.7 macrophages and endotoxin induced shock model in mice. J. Cell. Biochem., 117: 2327-2339.
- 84. Shen, K.H., A.C.H. Liao, J.H. Hung, W.J. Lee and K.C. Hu *et al.*, 2014. α -Solanine inhibits invasion of human prostate cancer cell by suppressing epithelial-mesenchymal transition and MMPs expression. Molecules, 19: 11896-11914.
- 85. Wang, L., Q.Q. Sun, S.J. Zhang, Y.W. Du and Y.Y. Wang *et al.*, 2016. Inhibitory effect of α -solanine on esophageal carcinoma *in vitro*. Exp. Ther. Med., 12: 1525-1530.
- Lee, K.R., N. Kozukue, J.S. Han, J.H. Park and E.Y. Chang et al., 2004. Glyco-alkaloids and metabolites inhibit the growth of human colon (HT29) and liver (HepG2) cancer cells. J. Agric. Food Chem., 52: 2832-2839.
- 87. Martuzzi, M., 2007. The precautionary principle: In action for public health. Occup. Environ. Med., 64: 569-570.